MYCOLOGIA

Vol. X

NOVEMBER, 1918

No. 6

DALDINIA VERNICOSA—A PYROXY-LOPHILOUS FUNGUS

ARTHUR S. RHOADS
(WITH PLATE 14)

The collector is thoroughly aware that numerous species of fungi occur prevailingly on burnt places. Although some of these forms are found elsewhere occasionally, many are so constantly associated with burnt places that they are sought only in such a habitat. In order to account for this peculiar association many plausible but inadequate reasons have been advanced. The fact remains, as stated by Seaver (1), that sterilization of the substratum by heat apparently brings about some change in the soil other than the simple elimination of competition in the destruction of bacteria and other fungi, which changes appear to be of vital importance in the cultivation of fungi which normally grow on a burnt substratum. Later experiments by Seaver and Clark (2), dealing with the artificial cultivation of a species of Pyronema, show that soil heated in various ways, especially by burning over the surface, becomes a very favorable nutrient medium for fungi of various kinds by reason of the large amount of food material rendered available through the heating of the materials in the soil. It is only natural to suppose that wood or bark, when burnt, likewise becomes a more favorable medium for the growth of certain fungi.

The writer has made several collections of *Daldinia vernicosa* (Schw.) Ces. & De Not. in various states throughout the East and generally finds it to be associated with fire-scorched trees. So far

[Mycologia for September (10: 231-275) was issued September 25, 1918.]

as can be determined this fungus is confined entirely to dicotyledonous trees but occurs upon a great number of species, preferably upon fire-scorched trunks. It attacks small saplings even more readily than larger trees and seems to prefer species of hickory for a host.

While making a survey of a burned area in the latter part of August, 1916, with a view to securing data in regard to the rapidity of the deterioration of standing fire-killed timber by one of our most common sap-rotting fungi, the writer was impressed by the abundance of Daldinia vernicosa on the dead trees. An area was selected near State College, Pa., that had been burned for the first time, a surface fire having swept through it I year and 3 months previously. As a result the trees on this area, with few exceptions, were scorched so badly that they were killed outright. From this burned area an average sample tract, 100 by 500 feet, was laid off and the following data were secured for each standing tree within this tract: species, diameter (measured to the nearest inch) at breast height, conditions (as to whether dead or living), and the species of fungi growing upon it as evidenced by the sporophores upon the trunk. The species of trees upon this tract in the order of their importance were white oak, scarlet oak, white pine, mocker nut hickory, red maple, chestnut, and pitch pine. The data obtained are given below, the trees being tabulated by diameter under each species:

Out of 71 scarlet oak trees upon this tract only 1 bore sporophores of *Daldinia vernicosa*. There also were present 15 red maples, 6 chestnuts, 37 white pines, and 1 pitch pine but no trees of these species bore sporophores. Out of a total of 363 dicotyledonous trees occurring upon this tract 46, or 13 per cent., bore sporophores of *Daldinia vernicosa* within 1 year and 3 months after the trees were scorched by fire. All the trees tabulated above, save one, were dead at the time of the reconnaissance.

The above figures clearly indicate how extensively and rapidly this ascomycete can propagate itself when afforded a favorable substratum. The accompanying photograph (Plate 14, A), taken in the latter part of August with a previous record of two months of dry weather, testifies to the luxuriant growth made by the

SUMMARY OF TREES BEARING SPOROPHORES OF DALDINIA VERNICOSA BY SPECIES AND SIZE

Species	D. B. H. inches	No, of trees on area	No. bearing sporophores of D. vernicosa	Per cent bear- ing sporophores of D. vernicosa
Quercus alba	. 2	12	3	25
64 49	. 3	51	10	31
44	. 4	37	5	14
44	. 5	40	5	12
46 44	. 6	33	6	18
# # #	. 7	27	1	4
24 44	. 8	19	0	0
86 68	. 9	8	0	0
** **	. 10	3	0	0
	. 11	6	0	0
Quercus alba, total trees		236	36	15
Hicoria alba	. 1	7	2	29
** **	. 2	12	4	33
61 41	. 3	6	2	33
44 44	. 4	5	0	0
86 88	. 5	3	0	0
44 18	. 6	1	0	0
" "	. 7	ī	1	100
Hicoria alba, total trees		35	9	26

sporophores of this fungus. Sporophores gathered and taken into the laboratory at this time shed copious quantities of spores.

Associated with Daldinia vernicosa was another pyrenomycete, Nummularia Bulliardi Tul. The stroma of this fungus is effused, thin, and crustaceous. It overspreads the surface of the inner bark, throwing off the epidermis for 6 inches or more in extent and is black and carbonaceous at maturity. This fungus, however, was found only on the white oak and scarlet oak trees. It is quite common throughout this region but always associated with dead oak trees. It is not, however, so restricted to fire-killed trees as is its associate, Daldinia vernicosa, but is apparently always associated with dead trees, preferably oak trees.

The genus *Daldinia* is characterized by the peculiar structure of the stroma, which is superficial, subglobose, and has a black and carbonaceous external layer when mature, in which the perithecia are imbedded. The stroma is softer inside, of a radiate-fibrous structure and concentrically zoned.

There are 24 species of Daldinia, mostly from the tropics, given

in Saccardo. For the most part they can be referred to Daldinia concentrica, which is a common and widely distributed plant occurring in almost every country in the world. In Australia this species assumes large size, frequently becoming two or three inches in diameter as it sometimes does in the western United States. In Europe, Daldinia durissima was proposed by Fries many years ago, but, according to Lloyd (3), no one else ever found it, a type at Kew being only the common D. concentrica. Massee found a specimen in tropical America (Trinidad) which he named Daldinia aspera. Lloyd, however, states that this specimen is not a Daldinia (3) at all but a Hypoxylon, probably H. cerebrinum (4). Léveillé discovered two species in the United States, D. cingulata and D. loculata, but Lloyd (3) states that both are the common D. concentrica. Two well-known and apparently distinct species of Daldinia occur within the United States, namely D. concentrica (Bolt.) Ces. & De Not. and D. vernicosa (Schw.) Ces. & De Not.

Peck (5), in his list of the plants of North Elba, reports Daldinia vernicosa on dead trunks of young, standing deciduous trees. He states that it is very doubtful if this and D. concentrica are really distinct species, and is of the opinion that connecting forms occur. It would appear that Peck had not collected Daldinia vernicosa as typified by the specimens in the Schweinitzian herbarium, for, if he had done so, it is difficult to see how such specimens could be considered identical with D. concentrica.

The stroma of *Daldinia concentrica* is subglobose or hemispheric, or rarely obovoid, while that of *D. vernicosa* is subturbinate and sometimes contracted behind into a thick stipe-like base which is often concentrically wrinkled. The stromata of both species become black when mature, but that of *D. vernicosa* becomes distinctly shining. When young and immature the stroma of *D. vernicosa* contains a large quantity of a colorless gelatinous substance which dries down at maturity, forming the radiate-fibrous substance between the concentric zones. At maturity practically all of the substance between the thin, blackish, concentric zones under the terminal, monostichous perithecial layer is made up of a colorless, radiate-fibrous, dry-gelatinous

substance. It is thus seen that the interior of the stroma of this plant is of a very heterogeneous texture. As a result of the loose texture of the radiate-fibrous inner substance the mature fruit-bodies can be crushed readily between the fingers. In *Daldinia concentrica* the interior of the stroma also is of a radiate-fibrous structure. Owing to its more homogeneous structure, however, it is fairly firm and solid, and specimens that have not been attacked by insects are very resistant to crushing. In the latter plant the radiate-fibrous substance is brown instead of colorless, as it is in



Fig. 1. Spores of Daldinia vernicosa showing various stages in the dehiscence of the exospore wall after treatment with dilute KOH; a, spore at time of shedding; b, spore showing the initial step in the dehiscence of the exospore wall; c and d, spores showing the casting off of the exospore membrane; e, cast-off exospore membranes, some with the valves still hinged together; f, a later stage of e, showing the return of the two valves to their original position. \times 500.

D. vernicosa (Plate 14, B), and the concentric zones are not so sharply defined as those of the latter species. As pointed out by Ellis and Everhart (6) the perithecia of D. concentrica are monostichous and not polystichous as stated by Saccardo. But little difference is exhibited by either the perithecia, asci, or spores of the respective species. The spores of D. concentrica are obliquely uniseriate with the ascus, inequilaterally elliptical, darkbrown, and finally opaque. They are somewhat variable in size but usually conform to $12.5-18\,\mu$ by $7-10\,\mu$. The spores of D. vernicosa are about the same size as in the preceding species but are somewhat smaller and less variable in size. They usually conform to the limits of $10-14.5\,\mu$ by $7-7.5\,\mu$.

The spores of Daldinia vernicosa are peculiar in that, when mounted in dilute (5 per cent.) KOH or NaOH, the exospore wall, which is colorless, quickly dehisces and separates from the spore, which is dark-brown. A single peripheral line of dehiscence occurs at the center of the spore and the two halves of the exospore wall usually break away from one another as two valves, or they may dehisce partially and bend backward as if they were hinged, thus allowing the spore to free itself from its peripheral membrane (Fig. 1). The spores of Daldinia concentrica also exhibit the same behavior, and, with equal facility. These observations on the dehiscence and shedding of the colorless exospore wall of these two species, when the spores have been mounted in dilute solutions of KOH and NaOH as well as certain other dilute alkaline solutions, have been confirmed by the careful and repeated examination of specimens from widely distant points in several localities. This dehiscence of the exospore wall is less evident, however, in old herbarium material.

Ellis and Everhart (6) sum up the differences between Daldinia vernicosa and D. concentrica as follows: "This (D. vernicosa) is distinguished from D. concentrica by its shining-black stroma, and the looser texture of the radiate-fibrous inner substance which is cut by 8-12 dark-colored, membranaceous horizontal plates or layers. These are very noticeable in a vertical section even in the young plant, while it is still covered with the conidial layer and before the terminal, subglobose, ascigerous stroma has begun to appear. In the mature state, the fibrous inner substance and the horizontal membranes disappear to a greater or less extent, and leave the stroma more or less hollow, so that it may be easily crushed with the fingers, but in D. concentrica the inner substance remains firm and is also of a darker color."

Daldinia concentrica, according to Lindau (7), is of cosmopolitan occurrence on dicotyledonous wood, while D. vernicosa, according to Saccardo (8), is less widespread in its distribution. In addition, the latter species generally occurs on burned woody stems, whereas the former species does not seem to be pyroxylophilous.

It is often very difficult to secure mature specimens of D. con-

centrica, and sometimes exceedingly difficult to secure mature specimens of *D. vernicosa* that are free from insects. Even after excellent specimens are collected, the interior portions of the stroma usually are eaten out by the larvae that hatch out within the specimens, unless they are quickly oven-dried.

In addition to his own collections the writer has examined specimens of both plants in the herbaria of Dr. L. O. Overholts, The Pennsylvania State College, The New York State College of Forestry, Office of Pathological Collections in the U. S. Bureau of Plant Industry, the Schweinitzian herbarium in the Academy of Natural Sciences at Philadelphia, and the collections of the Office of Investigations in Forest Pathology. The Schweinitzian herbarium contains the type specimens of Daldinia vernicosa, which were first described as Sphaeria vernicosa by Schweinitz from specimens collected at Salem, North Carolina.

SUMMARY

I. Daldinia vernicosa, as is typical of certain other fungi, occurs prevailingly on a substratum of burnt wood, and is to be regarded as a pyroxylophilous fungus.

2. In its occurrence, it apparently is confined to dicotyledonous species and attacks fire-killed saplings, particularly those of hick-ory, with great vigor.

3. Out of a total of 363 dicotyledonous trees occurring upon an average sample tract (100 by 500 feet) of a burned area, 46, or 13 per cent., bore sporophores of *Daldinia vernicosa* within 1 year and 3 months after the trees were scorched by fire.

4. Of the 24 (mostly tropical) species of *Daldinia* given in Saccardo, most of them can be considered as mere growth forms or ecological expressions of *Daldinia concentrica*, a widely distributed plant of cosmopolitan occurrence.

5. Only two species of *Daldinia* occur in the United States, *D. concentrica* and *D. vernicosa*, which appear to be morphologically quite distinct.

6. The dehiscence of the colorless exospore wall occurs along a single central peripheral line and seems to be a characteristic feature of regular occurrence with the spores of both *Daldinia*

vernicosa and D. concentrica, when mounted in dilute alkaline solutions.

Office of Investigations in Forest Pathology, Bureau of Plant Industry, Washington, D. C.

BIBLIOGRAPHY

- Seaver, Fred J. Studies in pyrophilous fungi—I. The occurrence and cultivation of Pyronema. Mycologia 1: 131-139. pls. 9-12. 1909.
- Seaver, Fred J., and Clark, Ernest D. Studies in pyrophilous fungi—II.
 Changes brought about by the heating of soils and their relation to the growth of Pyronema and other fungi. Mycologia 2: 109-124. pls. 24-26. 1910.
- 3. Lloyd, C. G. Daldinia vernicosa. In Mycological notes. No. 43, p. 604.
- Lloyd, C. G. Hypoxylon cerebrinum. In Mycological notes. No. 42, p. 579. 1916.
- Peck, C. H. Plants of North Elba, Essex Co., N. Y. New York State Museum Bul. 6: 67-266. 1899. (See p. 229.)
- Ellis, J. B., and Everhart, B. M. Daldinia-vernicosa. In The North American Pyrenomycetes, p. 661. 1892.
- Lindau, G. Daldinia DeNot. In Engler and Prantl's Die natürlichen Pflanzenfamilien. Teil I, Abteilung I, p. 487. 1897.
- 8. Saccardo, P. A. Daldinia vernicosa. In Sylloge. Vol. 1, p. 394. 1882.

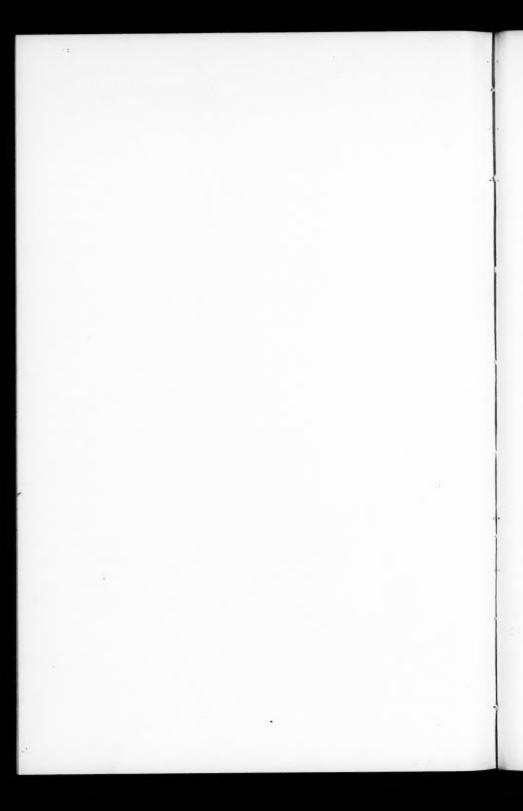
EXPLANATION OF PLATE 14

A. Trunk of white oak (Quercus alba L.) one year and three months after it was killed by a light surface fire, showing the abundance of Dalidinia vernicosa. The trunk bears a 6-inch rule.

B. Sporophores of *Daldinia vernicosa*, showing both external and sectional views, natural size.



DALDINIA VERNICOSA (Schw.) Ces. and De Not.



NEW JAPANESE FUNGI NOTES AND TRANSLATIONS—V

Työzaburó Tanaka

Physalospora minuta I. Miyake sp. nov. in Sangyô Shikenjô Hôkoku (Technical Report of the Imperial Sericultural Experiment Station), Tôkyô, Japan. 15: 314, pl. 16, figs. 1-3, T. 5, xii, Dec. 1916. (Japanese.)

Perithecia sunk in the matrix, with ostiola erumpent, ellipsoid or globoid, $150-200\,\mu$ in diam. and nearly $125\,\mu$ in height; perithecial wall black, pseudo-parenchymatous; ostiola $45-50\,\mu$ long, about $40\,\mu$ across; asci clavate-cylindric, thick-walled above, $60-70\times13-18\,\mu$, paraphysate, octosporous; paraphyses filiform, abundant, forming thick periphysatic tissue; ascospores subbiseriate, fusoid or ellipsoid, obtuse, minutely granulate, $18-22\times8-9\,\mu$.

On living twigs of Morus alba.

Type localities: Gifu-ken, Kaidzu-gun, Shiroyama-mura, Mar. 1909, I. Miyake; Fukui-ken, Mar. 1909, K. Hara; Kyôto-fu, Ayabe-chô, Apr. 26, 1915, I. Miyake.

Illustrations: Three lithographic figures showing detailed structure of the fungus.

Mostly appears in presence of *Macrophoma minuta* Berl. the pycnidia of which are surrounded by blackened hyphae commonly known as subiculum, which occur simultaneously with the formation of perithecia of the present species. Symptoms of the two are identical, shown by minute, gregarious, elevated spots covering certain areas of the twigs. As to the evidence of genetic relationship between the two, since no ascogenous form of the former species has been reported, the new name is given as above.

ASCOCHYTA MORI I. Miyake sp. nov. in Sangyô Shikenjô Hôkoku (Imperial Sericultural Experiment Station, Technical Report), 15: 345, Pl. 17, figs. 17–18, T. 5, xii, Dec. 1916. (Japanese.) Pycnidia ellipsoid or conoid, immersed, later erumpent, with single, apical ostiolum, $160\,\mu$ across; ostiola papillate, dark-bordered; perithecial wall pseudo-parenchymatous, not very thick, paler inside, increasingly dark outwards; mycelia surrounding perithecial wall dark-colored, mixing with colorless ones which predominate farthest from pycnidia; pycnospores mostly elliptic, frequently cylindric with blunt ends, or ovoid, septate at the middle, not constricted, 9–11 \times 3.5–40 μ , walls colorless, protoplasm pale-greenish, usually not conspicuously granulate but rarely one-nucleate in each cell; pedicel colorless and hyaline, short.

On branches of Morus alba.

Type localities: Fukui-ken prefecture, Japan, March, 1909, K. Hara; Idu-no-kuni, Shidzuoka-ken, Japan, Apr., 1909, I. Miyake.

Illustrations: Two black and white lithographic figures showing pycnidium and pycnospores.

Ascochyta moricola Berl. differs from this species in having dark-colored fusoid pycnospores pointed at both ends, and constricted at the septum.

Note: As the name Ascochyta mori has already been used by R. Maire (Ann. Myc. 114: 354, Aug. 1913), I propose a new name, Ascochyta Miyakei for this species.

Stagnospora mori I. Miyake sp. nov. in Sangyô Shikenjô Hôkoku

(Technical Report, Imperial Sericultural Experiment Station), Tôkyô, Japan. 15: 348, pl. 17, figs. 22, 23. T. 4, xii, Dec. 1916. Japanese.)

Pycnidia sub-epidermal, walls of thick pseudo-parenchymatous tissue, dark-brown, ellipsoid or globoid, erumpent with short papilliform openings, $130-160 \times 120-150\,\mu$; ostiola black and darker than the pycnidial wall; pycnospores cylindric, slightly curved, sometimes inequilateral, rounded at both ends, 3-septate, one septum formed earlier, more or less constricted, colorless, hyaline, granulate, germinating from either end or from both at the same time; $21-26\times6-9\,\mu$; pedicel short, small; paraphyses filiform, straight or slightly curved and twisted, the innermost the longest, shortening toward the opening.

Illustrations: Two black and white lithographic figures showing pycnidium and pycnospores.

On twigs of Morus alba.

Type locality: Yamagata-ken (prefecture) Yonezawa-shi, Mar. 1915, I. Miyake.

It is often observed that the fungus causes the host tissues to disintegrate and usually only bast fibers are left unattacked.

ROBILLARDA MORI I. Miyake sp. nov. in Sangyô Shikenjô Hôkoku (Technical Report of the Imperial Sericultural Experiment Station), 15: 346, pl. 17, fig. 19. T. 4, xii, Dec. 1916. (Japanese.)

Pycnidia hypo-epidermal, later erumpent with a single ostiolum, black, globoid or ellipsoid, $200\,\mu$ across; ostiola papillate, short and small; pycnospores cylindric, $15-18\times2.5-3\,\mu$; more or less thickened at the middle portion, slightly rounded at the base, and rather pointed at the apex, straight or slightly curved, colorless to pale-greenish, septate at the middle, not constricted, with 3-4 bristles at the end; bristles equal in length.

On dead branches of Morus alba (rare).

Type locality: Fukui-ken prefecture, Japan, March, 1909, K. Hara.

Differs from R. Cavarae Tognin, which has pycnospores with long pedicels measuring 40-50 μ ; and from R. Celtidis Scalia, characterized by having paraphyses 40-45 μ long.

Cytodiplospora Mori I. Miyake, sp. nov. in Sangyô Shikenjô Hôkoku (Technical Report, Imperial Sericultural Experiment Station), Tôkyô, Japan. 15: 347, pl. 17, figs. 20–21. T. 5, xii, Dec. 1916. (Japanese.)

Stromata scattered or gregarious, black, hemispherically elevated above, then disclosed, rupturing the epidermis, $\frac{1}{2}-\frac{2}{3}$ mm. in diam., round or ellipsoid, pseudo-parenchymatous; pycnidia 4–5, sometimes more than 10 in one stroma, globoid or ellipsoid, with short, flat ostiola; pycnidial wall made up of finely and densely fascicled hyphae, colorless inside; pycnospores colorless, hyaline or pale-greenish, guttulate, cylindric with round ends, ellipsoid or ovoid, even, sometimes irregular, straight or curved, uniseptate, septa centric or eccentric, constricted or not constricted, variable in size, 6–15 \times 3–5 μ .

On living twigs of Morus alba.

Type locality: Tôkyô-fu (prefecture) Nakano-chô, May, 1915, I. Miyake.

Illustrations: Two black and white lithographic figures showing pycnidia and pycnospores.

Found nowhere else, parasitic; mycelium intercellular and with haustoria entering the host cells.

DIMEROSPORIUM MORI Y. Endô sp. nov. in Dainippon Sanshi Kwaishô (Journal of the Sericultural Association of Japan), **26**³⁰³: 300, fig. B on p. 288, Apr. 1, 1917. (Japanese.)

Perithecia large, ellipsoid, 110–120 μ high, 130–140 μ across, without appendages, dark-brown; perithecial wall consisting of large cells containing several oil globules in each cell; asci numerous, clavate, thin-walled, $60–70\times12–15\,\mu$, 8-spored; ascospores almost definitely biseriate, oblong, subacute at both ends, $7–8\times5–7\,\mu$, yellowish-brown, uniseptate, with 1–2 shining oil globules in each cell.

Epiphytic on leaves of *Morus alba* (mostly on variety *Nezumi-gaeshi*), occurring with a species of *Meliola*. Catenulate hyphae, unicellular microconidia, multicellular macroconidia, gemmae, spermogonia, and pycnidia were observed, but it was not determined to which species they belong.

Locality: Ueda, Chiisagata-gun, Nagano-ken, Japan, nursery ground of Ueda Sericultural College, and mulberry fields of Tokida section east of the college grounds.

BUREAU OF PLANT INDUSTRY, WASHINGTON, D. C.

NOTES AND BRIEF ARTICLES

Professor F. S. Earle spent some days at the Garden about the middle of August and then sailed for Porto Rico, where he will investigate for the United States Government a serious and rather obscure disease of sugarcane.

Mr. Stephen C. Bruner, formerly assistant pathologist at the Estación Experimental Agronómica, Santiago de las Vegas, Cuba, has been appointed pathologist to succeed Mr. John R. Johnston, now head of the Office of Sanadad Vegetal, Havana.

Venenarius pantherinoides, described by Murrill from Seattle in 1912, has recently been collected at Olympia, Wash., by Miss M. McKenny, who states that it was eaten by two persons with almost fatal results.

Mr. F. W. Haasis reports in the *Journal of Agricultural Research* for 1917 that young pines in plantations at Portland, Conn., were found to be dying around ant-hills, the trouble being usually associated with fungous and scolytid infestations. Ants are thought to be instrumental in spreading the disease.

In a recent number of *Science*, Professor Gage suggests an excellent method for the preparation of lantern slides showing diagrams, tables, etc. This consists in first covering the glass with a thin coating of varnish and then drawing upon it with a pen, using India ink. Such slides may be covered and bound if desired for permanent use.

An unprecedented danger from fire in the National Forests of the Northwest and Pacific Coast, owing to early drought, high winds, electrical storms, labor shortage, and depletion of the regular protective forces because of the war, has made necessary a loan of \$1,000,000 to the Forest Service from the President's special defense fund.

A particularly large and excellent collection of fungi, accompanied by beautiful photographs and many notes, has been sent to the Garden for determination by Henry J. Rust, of Coeur d'Alene, Idaho. This region is interesting because it lies near the boundary line between the Rocky Mountain region and the Pacific coast.

Several wood-destroying fungi have been recently sent in for determination by Professor R. J. Blair, of McGill University, Montreal; among them Coriolus pubescens, Coriolellus serialis, Gloeophyllum trabeum, Lentodium tigrinum, Pyropolyporus conchatus, Phaeolus sistotremoides, and Micromphale ulmarium. Specimens of Lentodium tigrinum are particularly well developed, which is rather rare for this species.

A gigantic specimen of Ganoderma sessile Murrill, a bracket fungus with a reddish, shining surface, was brought to the Garden early in September by Mr. Michael Dougherty, who found it at the base of a dead red maple in Central Park. The specimen in its dried state measured 18 inches across and consisted of several layers superimposed, making the entire cluster about 6 inches thick. This species has been said by some to be identical with Fomes lucidus of Europe, but it is quite certain that no European mycologist would recognize it in this New York form.

A serious disease of wheat, long known in Europe, has recently been found in certain parts of the United States, particularly in Virginia, where in some fields losses have been as high as 40 per cent. of the crop. The disease, caused by a small nematode, or eelworm, usually affects the wheat heads, although it may occur on all parts of the plant above ground. Affected heads stay green and ripen late and are smaller than those not affected. The chaff usually opens at a wide angle. In place of grains of wheat,

the affected heads contain dark, hard galls somewhat shorter and thicker than wheat grains. Control measures consist of planting only disease-free seed, practicing crop rotations, and preventing the spread of the nematodes from one field to another by means of infected soil which may cling to the feet of men or animals or to farm implements.

A recent paper by Stakman and others, in the Journal of Agricultural Research, treats of the impossibility of breeding cereals permanently resistant to rust. The facts recorded in the paper, supported by experimental work in the rust nursery and by field observations, indicate that rust resistance is comparable with other permanent characters, and that it is not primarily controlled by seasonal conditions, soil type, geographical location, or other cultural conditions. It is rather an hereditary character, which cannot be produced by the accumulation of fluctuating variations within a susceptible line, nor broken down by changes in the host or parasite. The resistance of wheat varieties may vary in different regions because of the presence of different biological forms of rust.

Mr. Frank N. Meyer, one of the most successful agricultural explorers ever employed by our Government, was missed from a steamer on the Yangste River early in June and his body was afterwards recovered. There were no indications as to the cause of death. Many duplicates of Mr. Meyer's collections of fungi in the Orient came to the Garden for determination and were deposited in the herbarium. Only recently, Mr. Meyer succeeded in discovering the chestnut canker on wild chestnut trees in China, the original home of the disease.

It is stated by Mr. J. B. Rorer that an alga, Cephaleuros virescens, causes a leaf-fall and die-back disease of cacao on practically every estate in Trinidad. This disease has been under observation since 1912 and has been described as attacking any cacao tree at any time during the year, but more readily during the last

two months of the dry season, especially if the trees are not in a good situation or condition. The disease has been called die-back and sun-scald, but the author suggests the name of algal disease in order to distinguish it from true die-back and sun-scald, which are said to be caused by a Diplodia. Spraying with Bordeaux mixture has been attended by beneficial results, and attention to tillage, drainage, shade, and protection from wind are also considered essential to the complete control of the disease.

In the Journal of Agricultural Research for 1918, W. H. Long and H. M. Harsch describe a method for differentiating various wood-rotting fungi by their cultural characters alone when grown upon artificial media. It is claimed that when cultural characters of closely related but really distinct species are compared, marked and constant differences in the character of the mycelium will be found on certain corresponding agars in the series of cultures representing the two species, while if the fungi are really of the same species, no constant differences will occur. Basing the conclusion on these facts, the authors state that unknown rots can be identified by making pure cultures of the causative organisms from diseased wood.

Professor Bruce Fink, of Miami University, has contributed the following note:

"On the fifth of September, 1918, I was called to examine what a farmer had brought to Oxford, Ohio, and was exhibiting as an unusual mushroom. I found the exhibit to be a cluster of Clitocybe illudens, 90 inches in circumference, 15 inches high, and 44 inches from one side over the top to the opposite side. The cluster was compressed-hemispheric in form. There were approximately 300 plants that stood out so that they could be seen readily, and some bystander thought there were as many as 400 in all, counting those that were compressed between the ones that were plainly visible. Seeing this unusual cluster of fungi recalls that in 1896, I found at Fayette, Iowa, a specimen of Lycoperdon giganteum which was 85 inches in circumference. The plant was of the usual form for this species, and was, as I recall, between

18 and 24 inches high. Unfortunately, I took only the measurement of the circumference. The plant would sit on top of a bushel-and-a-half basket of the usual form and extend beyond the basket on all sides."

In order to prevent a large percentage of loss in the new crop of potatoes after storing, the Department of Agriculture is making the following suggestions to farmers:

Get rid of every bit of vegetable matter in the storage cellar; sweep and brush until it is clean; then give a thorough dose of fungicide, either gas or spray, the quickest and easiest to apply being formaldehyde gas. For each 1,000 cubic feet of space, use 10 ounces of formaldehyde and 5 of potassium permanganate. Pour the formaldehyde over the permanganate in a deep container, and then leave the cellar immediately, because the gas is given off at once. Should it be found that these chemicals are too expensive, the Department recommends Bordeaux mixture of 5-5-50 strength. It may be applied with hand sprayer, pump, or broom; it is effective when thoroughly used and it does not cost much. It is expecting too much, says the department, to look for potatoes fit for market from a dirty, ill-ventilated cellar. Time, money, and work spent in growing a crop are wasted if the potatoes are stored where dead potatoes are carrying over the organisms that cause rots. Dry-rot attacks newly stored potatoes through bruises and wounds and spreads throughout the stored supply. Many farmers have cellars that now contain piles of sacks of potatoes, all rotten, sacks and all, and constituting a wet, foul mass that helps to decay the timbers and menaces the crop to be stored.

Byron David Halsted

Professor Halsted died at his home in New Brunswick, New Jersey, on August 28, 1918, after a protracted illness. He had occupied the Chair of Botany in Rutgers College for nearly thirty years and had previously been professor in the Iowa State University.

Professor Halsted has served as a member of the Advisory Board of "North American Flora," published by the New York Botanical Garden, since the commencement of that work in 1905; and, during the several years preceding, while it was in the organization stage, he was an active member of the group of American botanists who made the enterprise possible. He has been president of the Society for the Improvement of Agricultural Science and of the Botanical Society of America; edited the American Agriculturist for a period; and has also been one of the editors of the Torrey Botanical Club.

He was a highly successful and greatly beloved teacher and investigator of renown. His most important publications have been in the fields of agricultural botany and plant diseases, and they include over 300 titles. His loss is a deep personal bereavement to his many friends and professional associates.

INDEX TO AMERICAN MYCOLOGICAL LITERATURE

- Adams, J. F. Origin and development of the lamellae in Schizophyllum commune. Mem. Torrey Club 17:326-333. pl. 9.
 f. a, b. 10 Je 1918.
- Allard, H. A. Effects of various salts, acids, germicides, etc., upon the infectivity of the virus causing the mosaic disease of tobacco. Jour. Agr. Research 13:619-637. 17 Je 1918.
- Arthur, J. C., & Johnston, J. R. Uredinales of Cuba. Mem. Torrey Club 17:97-175. 10 Je 1918. [Illust.]

Includes new species in Cronartium (1), Cionothrix (1), Ravenelia (1), Uromycladium (1), Puccinia (3), Aecidium (2), and Uredo (3).

- Atkinson, G. F. Six misunderstood species of *Amanita*. Mem. Torrey Club 17:246-252. 10 Je 1918.
- Blodgett, F. H. Weather conditions and crop diseases in Texas.

 Mem. Torrey Club 17:74-78. 10 Je 1918.
- Brown, N. A. Some bacterial diseases of lettuce. Jour. Agr. Research 13: 367-388. pl. E., 29-41. 13 My 1918.
- Burlingham, G. S. A preliminary report on the Russulae of Long Island. Mem. Torrey Club 17: 301-306. 10 Je 1918.
- Carpenter, C. W. A new disease of the Irish potato. Phytopathology 8:286, 287. pl. 1. Je 1918.
- Conn, H. J. The microscopic study of bacteria and fungi in soil. New York Agr. Exp. Sta. Tech. Bull. 64: 1-20. Ja 1918.
- Cook, M. T. Common diseases of berries. New Jersey Agr. Exp. Sta. Circ. 88: 1-11. f. 1-6. 1 D 1917.
- Cook, M. T. Common diseases of garden vegetables and truck crops. New Jersey Agr. Exp. Sta. Circ. 89: 1-22. f. 1-12. 1 D 1917.
- Coons, G. H. Michigan potato diseases. Michigan Agr. Exp. Sta. Spec. Bull. 85: 1-49. f. 1-41. Mr 1918.

- Cooper, J. R. Studies of the etiology and control of blister canker on apple trees. Nebraska Agr. Exp. Sta. Research. Bull. 12: 1–117. pl. 1–24. 15 D 1917.
- Davis, J. J. Tilletia on wheat in North Dakota. Phytopathology 8:247. My 1918.
- Dodge, B. O. Studies in the genus Gymnosporangium—I. Notes on the distribution of the mycelium, buffer cells, and the germination of the aecidiospore. Brooklyn Bot. Gard. Mem. 1: 128-140. pl. 1+f. 1-5. 6 Jl 1918.
- Dodge, B. O., & Adams, J. F. Some observations on the development of *Peridermium Cerebrum*. Mem. Torrey Club 17:253-261. pl. 4-6+f. 1-3. 10 Je 1918.
- **Duff, G. H.** Some factors affecting viability of the urediniospores of *Cronartium ribicola*. Phytopathology 8:289–292. f. I. Je 1918.
- Fitzpatrick, H. M. The life history and parasitism of Eocronartium muscicola. Phytopathology 8:197-218. pl. 1+f. 1-4. My 1918.
- Fracker, S. B. Effect of crown gall on apple nursery stock. Phytopathology 8:247. My 1918.
- Gillespie, L. J. The growth of the potato scab organism at various hydrogen ion concentrations as related to the comparative freedom of acid soils from the potato scab. Phytopathology 8:257-269. f. i. Je 1918.
- Gilbert, W. W., & Gardner, M. W. Seed treatment control and overwintering of cucumber angular leaf-spot. Phytopatholgy 8:229-233. My 1918.
- Graff, P. W. Philippine micromycetous fungi. Mem. Torrey Club. 17:56-73. 10 Je 1918.
- Ascophanus verrucosporus, Meliola Litseae, Phyllosticta Brideliae and Actinothyrium Hopeae, spp. nov. are described.
- **Keitt, G. W.** Inoculation experiments with species of *Coccomyces* from stone fruits. Jour. Agr. Research 13:539-570. pl. 55-59 + f. 1-3. 10 Je 1918.
- **Levine, M.** The physiological properties of two species of poisonous mushrooms. Mem. Torrey Club 17:176-201. pl. 1, 2 + f. 1, 2. 10 Je 1918.

- Lyman, G. R. The relation of phytopathologists to plant disease survey work. Phytopathology 8:219-228. My 1918.
- Martin, G. W. Brown blotch of the Kieffer pear. Phytopathology 8:234-238. f. 1-9. My 1918.
- McClintock, J. A., & Smith, L. B. True nature of spinachblight and relation of insects to its transmission. Jour. Agr. Research 14: 1-60. pl. A, I-II + f. I. I Jl 1918.
- McCubbin, W. A. Public school survey for currant rust. Phytopathology 8:294-297. Je 1918.
- McDougall, W. B. Some edible and poisonous mushrooms. Bull. Illinois State Lab. Nat. Hist. 11:413-555. pl. 86-143+f. 1. N 1917.
- Muncie, J. H. Experiments on the control of bean anthracnose and bean blight. New York Agr. Exp. Sta. Tech. Bull. 38: 1-50. pl. 1-4. D 1917.
- Murrill, W. A. Collecting fungi at Delaware Water Gap. Mem. Torrey Club 17:48-51. 10 Je 1918.
- Nishimura, M. A carrier of the mosaic disease. Bull. Torrey Club 45: 219-233. pl. 7. 20 Je 1918.
- Olive, E. W. Potato diseases. Brooklyn Bot. Gard. Leaflets 6: [1-4]. 29 My 1918.
- Orton, W. A. Breeding for disease resistance in plants. Am. Jour. Bot. 5:279-283. Je 1918.
- Pammel, L. H. The extermination of the common barberry to prevent crop leakage due to stem rust. Iowa Conservation 2: 4-8. Mr 1918. [Illust.]
- Pierce, R. G. Notes on Peridermiums from Ohio. Need of pathological viewpoint in nursery inspection. Phytopathology 8:292-294. Je 1918.
- Potter, A. A. The effect of disinfection on the germination of cereal seed. Phytopathology 8:248, 249. My 1918.
- Rands, R. D. Early blight of potato and related plants. Wisconsin Agr. Exp. Sta. Research Bull. 42: 1-48. f. 1-10. Ap 1917.

- Rosenbaum, J., & Ramsey, G. B. Influence of temperature and precipitation on the blackleg of potato. Jour. Agr. Research 13:507-513. 3 Je 1918.
- Seaver, F. J., & Horne, W. T. Life-history studies in Sclerotinia.

 Mem. Torrey Club 17: 202-206. pl. 3. 10 Je 1918.

 Sclerotinia Geranii sp. nov. is described.
- Smith, E. F., Godfrey, G. H. Brown rot of Solanaceae on Ricinus. Science II. 48: 42, 43. 12 Jl 1918.
- Steinberg, R. A. A study of some factors influencing the stimulative action of zinc sulphate on the growth of *Aspergillus niger*, I. The effect of the presence of zinc in the cultural flasks. Mem. Torrey Club 17:287-293. 10 Je 1918.
- Stevens, H. E. Lightning injury to citrus trees in Florida. Phytopathology 8: 283–285. f. 1. Je 1918.
- Stevens, N. E., & Wilcox, R. B. Further studies of the rots of strawberry fruits. U. S. Dept. Agr. Bull. 686: 1-14. 24 Je 1918.
- Stone, R. E. Common edible and poisonous mushrooms of Ontario. Ontario Dept. Agr. Bull. 263:1-24. Je 1918.
- **Taubenhaus, J. J.** Pox, or pit (soil rot), of the sweet potato. Jour. Agr. Research 13:437-450. pl. 51, 52. 27 My 1918.
- Weir, J. R. Experimental investigations on the genus Razoumof-skya. Bot. Gaz. 66: 1-31. f. 1-19. 15 Jl 1918.
- Weir, J. R. Forest disease surveys. U. S. Dept. Agr. Bull. 658: 1-23. f. 1-23. 12 Je 1918.
- Weldon, G. P. Pear growing in California. Month. Bull. State Comm. Hort. Calif. 7: 222-410. f. 1-186. My 1918. [Illust.] Contains chapter on diseases of pears.
- Young, V. H. Some factors affecting inulase formation in Aspergillus niger. Plant World 21:75-87. Ap 1918.

INDEX TO VOLUME X*

New names, and the final members of new combinations, are in bold face type.

Abies, 198; balsamea, 48; grandis, 9, 99, 100; lasiocarpa, 4, 99 Acacia Richii, 88 Acalypha virginica, 169 Acer neomexicanum, 241 Achillea lanulosa, 39 Aecidium Compositarium, 41; micropunctum, 37

Agaricaceae of tropical North America-VII, The, 15; VIII, 62 Agaricus, 16, 25, 29, 30, 67, 68, 69, 70, 81, 85; angustifolius, 73, 74; antillarum, 32; arvensis, 77; bambusigenus, 73, 74; bulbillosus, 22; bullaceus, 17; campanulatus, 31; campester, 73, 76, 77, 78; cinchonensis, 73, 76; commiscibilis, 19; coprinoceps, 21; dichrous, 30; disseminatus, 21, 26; Earlei, 74, 79; epibates, 19; euthugrammus, 21; fimicola, 32; fimetarius, 83; fortunatus, 33; guadelupensis, 74, 81; herradurensis, 74, 78; hiascens, 28; hololepis, 69; Hornei, 74, 80; je-junus, 74, 78; Johnstonii, 73, 75; minutulus. 26; modestus, ochraceidiscus, 74, 80; palmigena, 29; papilionaceus, 31, 32; phalenarum, 32; plumiger, 23; plutonius, 30; praemagnus, 74, 78; pratensis, 73, 77, 78; pseudotener, 25; Sallei, 81; scatigenus, 17; Shaferi, 74, 81; silvicola, 75; solidipes, 31; subpratensis, 74, 75; subsilvicola, 73, 75; subviridis, 30; tricholepis, 69; Venus, 73, 76; xuchilensis, 74, 79;

yucatanensis, 81 Agoseris, 242 Agrimonia striata, 41, 244, 254 Agropyron Bakeri, 258, 260; Smithii, 251, 262; tenerum, 37, 39, 251 Allantosporae, 240 Allionia linearis, 247, 248 Allodus commutata, 35; Douglasii, 35; vertisepta, 35

Notes on, 4 Amanita, 101; Frostiana, 48; muscaria, 48 Amanitopsis, 101; volvata, 48 Amarella heterosepala, 40; strictiflora, 40 Amorpha canescens, 260 Anastraphia, 167; Northrupiana, 166 Andropogon scoparius, 37, 38 Androsace diffusa, 242 Anellaria, 30 Anogra coronopifolia, 258, 260 Apiospora, 243; sepincoliformis, 243 Apiosporae, 243 Apiosporella, 243; cornina, 243 Arabis kochii, 198 Arbutus Menziesii, 100 Arenaria Fendleri, 39, 242, 247 Argentina anserina, 262; argentea, 262 Armillaria, 266; albolanatipes, 101; mellea, 10 Artemisia albula, 38; dracunculoides, 37; franserioides, 40; frigida, 240, 251, 261; gnaphalodes, 40; redolens, 37; scouleriana, 250 Arthrobotryum (?) pestalozzioides, Arthur, J. C., Uredinales of Costa Rica based on collections by E. W. D. Holway, 111 Asclepias speciosa, 254 Ascochyta Boutelouae, 257; gramini-

Alnus tenuifolia, 239, 241

Altitudinal range of forest fungi,

cola, 257; Miyakei, 286; mori, 285, 286; moricola, 286
Ascochytula, 258; agropyrina, 258
Ascomycetes, 239
Ascomycetes and lower fungi from New Mexico, New or noteworthy, 239
Aster Novae-Angliae, 254; vallicola, 256; Wootonii, 37
Asterophora Clavus, 97

* It has been considered unnecessary to include here the species listed in the three following articles, since they are already indexed or specially listed.

Arthur: Uredinales of Costa Rica. See p. 150. Brenckle: North Dakota fungi. See p. 199. Weir: Altitudinal range of forest fungi. See p. 11. Astragalus oreophilus, 254 Atriplex canescens, 240, 261 Atylospora, 15, 18, 21, 29; albipes, 18, 22, 33; bulbillosa, 18, 22; byssina, 18, 20, 33; cinchonensis, 19, 24, 33; commiscibilis, 18, 19; coprinoceps, 18, 21; cubensis, 18, 23, 33; diminutiva, 18, 19, 33; epibates, 18, 20; euthugramma, 18, 21; fuliginosa, 18, 25, 33; lateritia, 18,20, 33; mammillata, 19, 23, 33; mexicana, 18, 21, 33; Musae, 18, 21; pallidispora, 18, 22, 33; plana, 19, 24, 33; plumigera, 18, 23; pseudotenera, 19, 25; Roystoniae, 19, 24; tigrina, 18, 19 Azalea mollis, 164

Baccharis Wrightii, 248 Bacterium Juglandis, 99 Betula fontinalis, 251; occidentalis, 100 Bjerkandera adusta, 109 Blepharoneuron tricholepis, 251 Boletus luteus, 45 Botrytis, 86; canescens, 86; liliorum, Bouteloua gracilis, 257 Brenckle, J. F., North Dakota fungi

-II, 199 Bromus ciliatus, 37; Porteri, 37 Bruner, Stephen C., Johnston, John R., and, A Phyllachora of the royal

Burlingham, Gertrude S., New species of Russula from Massachusetts, 93

Calamagrostis, 167; hyperborea americana, 252

Camarosporium Amorphae, 260; compositarum, 261; Estrelti, 261; patagonicum, 261; wistarianum, 165; yuccaesedum, 261

Campanularius, 16, 29, 30; anomalus, 31, 32; campanulatus, 31, 32; solidipes, 31, 32, 33

Carex, 167; Douglasii, 40; nebraskensis, 38

Cassiope mertensiana, 4 Castilleja integra, 36, 263; linariaefolia, 36; sulphurae, 36, 37 Cathartolinum australe, 36 Cenangium, 46; urceolatum, 46

Cephaleuros virescens, 291 Cercocarpus montanus, 240, 250, 252 Cercospora montana, 263; pini-densi-

florae, 89 Ceriospora Dubyi, 244; montaniensis, 244, 246

Cerrena unicolor, 110 Chalymota, 30 Chamaecyparis, 189

Chamaesyce serpyllifolia, 41 Cheirinia, 262 Chenopodium album, 259 Chitonia, 85 Chrysomyxa Weirii, 98 Chrysopsis hispida, 247 Chrysothamnus graveolens, 247 Cirsium coloradense, 37; ochrocentrum, 37; polyanthus, 41; Richardsoni, 41

Cladosporium fasciculatum, 263: herbarum, 262; herbarum cerealium, 263

Clarkeinda, 16

Citrus, 88

Clathrospora permunda, 248

Clavaria, 57; ardenia, 54, 55, 57; brachiata, 56; contorta, 53, 55, 56, 57; fistulosa, 53, 54, 55, 56, 57; inaequalis, 53; juncea, 53, 54, 56, 57; juncea vivipara, 56; macrorrhiza, 54, 55, 57; vermicularis, 53

Clavaria fistulosa group, The, 53 Clematis, 166; calycina, 166; integrifolia, 166; ligusticifolia, 38, 244, 251, 264; paniculata, 165, 166; vitalba, 166

Clitocybe farinacea, 180; illudens, 202

Coccomyces Kerriae, 99

Coleosanthus grandiflorus, 39; reniformis, 246

Colections by E. W. D. Holway Uredinales of Costa Rica based on, III

Collema, 236, 238; pulposum, 235, 236, 238

Collemaceae, A new genus and species of the, 235

Collemodes, 236; Bachmanianum, 237, 238 Collybia conigena, 55; esculenta, 55;

velutipes, 266 Comandra pallida, 254

Compositae, 247

Conidial formation in Sphaeronema fimbriatum, 155

Coniothyrium concentricum Yuccaglaucae, 257; myriocarpum, 257; olivaceum Salsolae, 257; olivaceum Thermopsidis, 257; sepium,

Convolvulus sepium, 256

Coprinopsis, 82

Coprinus, 19, 21, 28, 82, 84; armillatis, 82, 83; cinchonensis, 82, 85; cubensis, 82, 83; fimetarius, 82, 83; jalapensis, 82, 83; jamaicensis, 82, 84; mexicanus, 82, 84; mica-

ceus, 28, 83; plicatilis, 82, 85; Spraguei, 82, 85 Coriolellus serialis, 290 Coriolopsis rigida, 269 Coriolus prolificans, 110; pubescens, Cornus instolonea, 254, 255 Corticum, 239 Cortinarius, 15, 46; corrugatus, 234; salmonicolor, 46 Cortinopsis, 62 Costa Rica based on collections by E. W. D. Holway, Uredinales of, 111 Crataegus erythropoda, 36, 252, 253 16; alveolatus, Crepidotus. fumosifolius, 16; musicola, 16 Cronartium coleosporoides, 36, 37 Cryptostictis utensis, 260 Cucurbitaria Ribis, 250; Rosae, 250 Cultures with Melampsorae on Populus, 194 Cup-fungi, Photographs and descriptions of,-VII. The genus Underwoodia, 1 Cycloporus Greenei, 47 Cylindrocladium scoparium, 99 Cyrtorhynca ranunculina, 253 Cytodiplospora mori, 287 Cytospora chrysosperma, 266

Daedalea confragosa, 99; unicolor, Daldinia, 279, 280, 283, 284; aspera, 280; cingulata, 280; concentrica, 280, 281, 282, 283, 284; durissima. 280; loculata, 280; vernicosa, 277, 278, 279, 280, 281, 282, 283 Daldinia vernicosa-a pyroxylophilous fungus, 277 Danthonia, 252; intermedia, 248, 252 Dasiophora fruticosa, 36 Dasyschypha Willkommii, 9 Dasystephana Bigelovii, 38 Deconica, 15, 17; bullacea, 17, 18; scatigena, 17 Delphinium robustum, 245 Dematiaceae, 262 Dematium pullulans, 263 Descriptions of cup-fungi, Photographs and,-VII. The genus Underwoodia, 1 Deuteromycetes, 253 Diaporthe oligocarpoides, 244 Diatrype albopruinosa, 241; cornuta, 241; Standleyi, 240 Diatrypella Placenta, 241 Dictyochora Gambellii, 166 Didymella Eurotiae, 243; nigrescens, 242; nigrificans, 243 Didymosphaeriae, 244 Diervilla Diervilla, 164, 165 Dimerosporium mori, 288

Diplodia, 45; hyalospora, 165 Distribution of fungi in Porto Rico, The, 58 Dodge, B. O., Studies in the genus Gymnosporangium—III. The origin of the teleutospore, 182 Dothichiza populea, 170 Dothidea Haydeni, 256 Dothideaceae, 251 Dothideales, 243 Dothidella insculpta, 251 Dothiorella, 255; phomopsis, 255 Drosophila, 16, 18, 21, 62, 67; appendiculata, 62, 63, 64, 66, 71; atricastanea, 63, 66; brevipes, 62, 63; caespitosa, 63, 67; campestris, 63, 64; castaneidisca, 62, 63; flocculosa, 63, 64; jalapensis, 63, 65; pallidospora, 63, 64; tenuis, 63, 65; tepeitensis, 63, 65; truncatispora, 63, 66 Dugaldea Hoopesii, 41

Echinochloa zelayensis, 41
Echinodontium tinctorium, 5, 7, 10
Elymus canadensis, 38, 245
Entoloma, 30
Epilobium adenocladon, 263; novomexicanum, 41
Eriogonum alatum, 259; Jamesii, 40; racemosum, 40
Erysibaceae, 239
Eurotia lanata, 243, 254
Euryachora betulina, 251
Eutypella Brunaudiana
240; herbicola, 240

Fairman, Charles E., New or note-

Fabraea litigiosa, 253

worthy Ascomycetes and lower fungi from New Mexico, 239; Notes on new species of fungi from various localities-II, 164 Fink, Bruce, A new genus and species of the Collemaceae, 235; The distribution of fungi in Porto Rico, 58 Firmiana platanifolia, 90 Flocculosa, 233 Fomes annosus, 10; lucidus, 45, 290; officinalis, 269; pini, 5, 7, 10; ungulatus, 100 Forest fungi, Notes on the altitudinal range of, 4 Fungi from New Mexico, New or noteworthy Ascomycetes and lower, 239 Fungi from various localities-II, Notes on new species of, 164 Illustrations of-XXVIII, Fungi. 107; XXIX, 177

Fungi in Porto Rico, The distribution of, 58 Fungi-II, North Dakota, 199

Fungi, Notes and translations-, New Japanese, IV, 86; V, 285 Fungi, Notes on the altidudinal range

of forest, 4 Fungus, Daldinia vernicosa-a pyroxylophilous, 277

Fusarium, 267 Fusicladium Cerasi, 263 Fusicoccum putrefaciens, 46

Gaertneria acanthicarpa, 254 Galium boreale, 241, 258 Ganoderma sessile, 290 Gaultheria humifusa, 4 Gaura induta, 261, 262; parviflora, 261

Gautieria, 266; plumbea, 266 Genus and species of the Collemaceae, A new, 235

Genus Gymnosporangium-III, origin of the teleutospore, Studies in the, 182

Geophila, 70 Gibberidia arthrophyma, 246 Globifomes graveolens, 267 Glochidion obovatum, 88

Gleophyllum trabeum, 290 Gloeosporium Potentillae, 262; venetum, 46

Glyptosperma, 62 Gomphidius, 16, 69; jamaicensis, 69 Grindelia aphanactis, 41

Grossularia inermis, 35; leptantha, 241, 250 Gymnochilus, 62; caespitosus, 67;

campestris, 64; flocculosus, 64; Musae, 21; Roystoniae, 24 Gymnolomia multiflora, 37

Gymnopus, 177

Gymnosporangium, 182, 189, 191; Betheli, 36; clavariaeforme, 182, 183, 189, 190, 192; clavipes, 183, 190; fraternum, 183; globosum, 183, 187, 189, 192; juniperi-virginianae, 183; macropus, 182, 183, 184, 187, 191; nidus-avis, 190, 192; Sabinae, 182; transformans, 183

Gymnosporangium-III. The origin of the teleutospore, Studies in the genus, 182

Harper, Edward T., Hypoloma aggregatum and H. delineatum, 231; The Clavaria fistulosa group, 53 Hedysarum pabulare, 40 Helianthus annuus, 41, 261; Maxi-

miliani, 246, 251

Helicobasidium tanakae, 89, 90, 91 Heliopsis scabra, 245, 255

Helvella crispa, 2

Hendersonia Agropyri, 260; calycina, 166; Clematidis, 166; crastophila, 260; Eriogoni, 259; foliorum, 260; hortilecta, 165; Leucelenes, 249; Petalostemonis, 249, 260; Rubi Clematidis, 166; sarmentorum Clematidis, 166: Stanleyellae,

259; subcultriformis, 260 Heracleum lanatum, 254 Herpotrichia nigra, 6 Heuchera parvifolia, 39, 240 Hicoria alba, 279 Hieracium Fendleri, 38

Holway, Uredinales of Costa Rica based on collections by E. W. D.,

Hordeum trifurcatum, 42 Houstonia minor, 168

Hubert, Ernest E., Weir, James R., and, Cultures with Melampsorae on Populus, 194

Humulus americanus, 244, 259 Hura crepitans, 68 Hyalodidymae, 242, 257 Hyalophragmiae, 259 Hyalosporae, 241, 253 Hydnum, 110 Hymenochaete noxia, 45

Hypholoma, 16, 67, 68, 71, 84, 234; aggregatum, 231, 232, 233; aggregatum sericeum, 232, 233; bermudiense, 72; caespitosum, 67; campestre, 64; delineatum, 231, 232, 233, 234; echiniceps, 232, 233; fasciculare, 67, 68; flavovirens, 67, 68; flocculosum, 64; hypoxanthum, 232; lacrimabundum, 231, 232, 233, 234; papillatum, 67; populinum, 232, 233; Pseudostorea, 232, 233; rugocephalum, 234; silvestre, 233; Storea, 231, 232, 233; Storea caespitosum, 231, 232, 233; tuberculatum, 67, 68; velutinum, 232, 234

Hypholoma aggregatum and H. delineatum, 231 Hypholoma delineatum, Hypholoma

aggregatum and, 231 Hypholomopsis, 62 Hypocreaceae, 251 Hypoxylon cerebrinum, 280, 284

Hysteriaceae, 252 Hysterium Notarisianum, 252; Standleyanum, 252

Hysterographium Bakeri, 252

Illustrations of fungi-XXVIII, 107: XXIX, 177 Index to American mycological literature, 49, 102, 172, 226, 271, 295 Inocybe, 15, 222 Ionoxalis violacea, 41

Ipomoea batatas, 155 Iris missouriensis, 242 Irpex, 110 Isocoma heterophylla, 247, 254, 261

Japanese fungi, Notes and translations—, New, IV, 86; V, 285 Johnston, John R., and Bruner, Stephen C., A Phyllachora of the royal palm, 43 Juglans, 90 Juncus balticus, 263 Juniperus scopulorum, 36

Kalmia latifolia, 268 Kellermannia Sisyrinchii, 259; yuccaegena, 259 Kerria japonica, 90, 99 Koeleria cristata, 252, 263 Kuhnia rosmarinifolia, 38, 247

Laccaria amethystea, 178; laccata, 178, 179; striatula, 179
Lachnella flammea, 252; rhoina, 252
Lachrymaria, 62
Laciniaria punctata, 247
Lactaria Allardii, 265; coleopteris, 265; Curtisii, 265; furcatus, 265; lentus, 265; subplinthogalus, 265; subtorminosus, 265
Lactuca pulchella, 41, 262

Latix, 194, 195, 196, 198; europea, 197; laricina, 195; lyallii, 5, 197, 198; occidentalis, 194, 195, 196, 197
Lathyrus arizonicus, 40; decaphyllus, 40; leucanthus, 40

Lathyrus arizonicus, 40; decaphyllus, 40; leucanthus, 40
Lavauxia flava, 262
Ledum glandulosum, 4
Lehman, S. G., Conidial formation in Sphaeronema fimbriatum, 155
Lentinus lepideus, 8
Lentodium tigrinum, 290
Lenzites sepiaria, 8
Lepiota, 75, 81, 82, 83; cretacea, 82
Leptonia conica, 178
Leptoniella conica, 178
Leptosphaeria Coleosanthi, 246; cul-

Leptosphaeria Coleosanthi, 246; culmifraga minuscula, 245; Doliolum, 245; dumetorum, 244; Helianthi, 246; lupincola, 245; nigricans, 245; nigricans Grindeliae, 245; ogilviensis, 245; praeclara typhiseda, 245; Quamoclidii, 246; rubrotincta, 246; Senecionis, 248; tenera, 245
Leucelene arenosa, 249, 250

Ligusticum Porteri, 246 Ligusticum Porteri, 246 Lilium longiflorum, 86 Linum Lewisii, 36 Lithospermum multiflorum, 247 Locellina, 15 Lophidiopsis nuculoides, 252 Lophiosphaera perpusilla, 167 Lophiostoma collinum, 167; quadrinucleatum, 251 Lophiostomataceae. Lophiotrema Mo' m, 167; pusillum, 167; stenogramma, 167 Lophodermium arundinaceum, 252 Lower fungi from New Mexico, New or noteworthy Ascomycetes and,

239 Lupinus ingratus, 245 Lycoperdon giganteum, 292 Lycurus phleoides, 248 Lygodesmia juncea, 262

Machaeranthera Bigeloviae, 245; Bigelowii, 263 Macrophoma cornina, 255; minuta, 285 Macrosporium commune, 263

Malvastrum coccineum, 39
Marasmius, 177; dichrous, 180; insititius, 181

Massachusetts, New species of Russula from, 93 Massaria, 240

Medicago sativa, 253, 254 Melampsora albertensis, 41, 194, 195, 196, 197, 198; Bigelovii, 36; Lini, 36; medusae, 194, 195, 196, 197, 198; occidentalis, 98

Melampsorae, 194, 198
Melampsorae on Populus, Cultures
with, 194
Melampsorella elatina, 198
Melanconiaceae, 262
Melanconiales, 262

Melanopsamma pomiformis, 244 Melanotus, 15, 16; fumosifolius, 16; musicola, 16

Melia Azedarach, 88 Melilotus alba, 244 Meliola, 288 Mertensia caelestina, 242, 248 Metasphaeria Senecionis, 248

Micranthes arguta, 39
Microdiplodia Anograe, 258; Diervillae, 165; galiicola, 241, 258;
Leucelenes, 249; Viciae, 258

Micromphale ulmarium, 290 Monarda comata, 38; stricta, 38 Moniliales, 262

Morus, 90; alba, 87, 91, 92, 285, 286, 287, 288; alba Nezumigaeshi, 288 Muhlenbergia cuspidata, 251; trifida, 30, 251

Murrill, W. A., Illustrations of fungi —XXVIII, 107; XXIX, 177; The Agaricaceae of Tropical North America—VII, 15; VIII, 62 Mycena galericulata, 179; viscidipes,

77

Mycosphaerella, 258; Iridis, 242; Oenotherae, 258; pachyasca, 242; Primulae, 242; tingens, 242 Myriangaceae, 46

Naematoloma, 67 Naucoria, 21; coprinoceps, 21; euthugramma, 21 Neopeckia coulteri, 6

New and noteworthy species, Studies in North American Peronosporales—VII, 168

New genus and species of the Collemaceae, A, 235

New Japanese fungi, Notes and translations—IV, 86; V, 285 New Mexico in 1916, Rusts and smuts collected in, 34

New Mexico, New or noteworthy Ascomycetes and lower fungi from,

New or noteworthy Ascomycetes and lower fungi from New Mexico, 239 New species of fungi from various localities—II, Notes on, 164

New species of Russula from Massachusetts, 93

North America—, The Agaricaceae of tropical, VII, 15; VIII, 62

North American Peronosporales— VII, New and noteworthy species, Studies in, 168

North Dakota fungi-II, 199

Nostoc, 235 Notes and brief articles, 45, 97, 170, 222, 265, 289

Notes and translations—, New Japanese fungi, IV, 86; V, 285 Notes on new species of fungi from

various localities—II, 164

Notes on the altitudinal range of forest fungi, 4

Noteworthy Ascomycetes and lower fungi from New Mexico, New or, 239

Noteworthy species, Studies in North American Peronosporales—VII, New and, 168

Nothopatella moricola, 91 Nummularia Bulliardi, 279 Nuttalia Rusbyi, 244

Odostemon repens, 41
Oenothera Hookeri, 262
Omphalia fibula, 179
Omphalopsis fibula, 179
Ophiobolus claviger, 250; collapsus, 250
Ophiodothis Haydeni, 256

Oreobatus deliciosus, 37 Origin of the teleuospore, Studies in the genus Gymnosporangium—III, The, 182 Osmunda cinnamomea, 94; regalis, 94 Otthia Clematidis, 244; fruticola, 244

Otthia Clematidis, 244; fruticola, 244 Oxytropis deflexa, 41; Lambertii, 262

Padus melanocarpa, 263 Palm, A Phyllachora of the royal, 43 Paneolus, 30; campanulatus, 31;

fimicola, 32; papilionaceus, 32; phalenarum, 32; solidipes, 31, 33 Panicum virgatum, 40

Patella oreophila, 253 Paulownia, 92; tomentosa, 90 Pedicularis fluviatilis, 38, 254; Grayi, 36

Pentstemon Torreyi, 37 Pericome caudata, 245

Peridermium coloradense, 198; ribicola, 35

Peronospora, 168, 169; grisea, 168; Polygoni, 168; Seymourii, 168 Peronosporales—VII, New and noteworthy species, Studies in North American, 168

American, 168 Pestalozzia, 264 Petalostemon oligophyllus, 249, 260 Pezizales, 252 Phaeoapiospora, 243

Phaeodictyae, 247, 260 Phaeodidymae, 244, 258 Phaeolus sistotremoides, 290

Phaeoapiosporae, 244

Phaeosporae, 241, 259 Phaeosporae, 241, 256 Philocopra coeruleotecta, 222

Phloeospora Oxytropidis, 262 Phlox, 35

Pholiota, 15, 71, 98; adiposa, 98; erebia, 234; flammans, 98; squarrosa, 98

Phoma. 241, 243, 245, 247, 250, 255, 266; asclepiadea, 254; Astragali, 254; complanata, 254; Corni-Sueciae, 254; Estrelti, 254; exigua, 254; herbarum, 250, 254; herbarum Medicaginis, 254; herbarum Solidaginis, 255; lupincola, 246; Mororum, 87; oleracea, 255; Rudbeckiae, 255; Sidalceae, 255; thalicinis

ictrina, 255; verbascicarpa, 164 Phomopsis ericaceana, 164; Kalmiae, 268

Photographs and descriptions of cupfungi—VII, The genus Underwoodia, 1

Phragmidium Andersoni, 36; imitans, 41; montivagum, 37; Peckianum, 37; Potentillae, 41 Phyllachora, 43; Ambrosiae, 251;

Blepharoneuri, 251; Haydeni, 256; Roystoneae, 43, 44; Trifolii, 251; vulgata, 251

Phyllachora of the royal palm, A, 43

Phyllactinia corylea, 239 Phyllodoce empetriformis, 4

Phyllosticta Crataegi, 253; Kuwacola, 87

Phyllostictaceae, 253

Physalospora Ambrosiae, 251; Arthuriana, 251; Galii, 241; minuta, 285

Picea engelmanni, 5, 198 Pilosace, 16, 68; hololepis, 69; tri-

cholepis, 69

Pinus albicaulis, 4; cembra, 5; contorta, 5; densiflora, 89; edulis, 35; flexilis, 5; pungens, 48; rigida, 48; Strobus, 48

Pittosporum undulatum, 90 Placosphaerella, 256

Placosphaeria decipiens, 256 Plasmopara Acalyphae, 169

Platystomum compressum, 252; phyllogenum, 166

Plectodiscella veneta, 46

Pleospora, 243; Bardanae, 247; coloradensis, 248; compositarum, 247; herbarum, 246, 247, 248; herbarum microspora, 247; infectoria, 248; rubicunda, 248; Senecionis, 248; vulgaris, 248; vulgatissima, 248

Pluteopsis, 18 Poa Bigelovii, 38; pratensis, 38

Poinsettia dentata, 41

Polemonium confertum, 248 Polygonum sawatchense, 254

Polyporus, 108; adustus, 109; albellus, 99; amorphus, 48, 109; brumalis, 108; elegans, 100; glomeratus, 170; irregularis, 109; leucospongia, 7, 8; Polyporus, 108; Schweinitzii, 10, 269

Polystictus conchifer, 108; hirsutus, 8 Polythrincium Trifolii, 263

Populus, 194, 198; angustifolia, 241, 244, 250, 257; aurea, 41; deltoides, 195; tremuloides, 194, 195, 196, 197; trichocarpa, 194, 195, 196, 197 Populus, Cultures with Melampsorae

on, 194

Poronidulus conchifer, 108 Porto Rico, The distribution of fungi

in, 58 Potentilla filipes, 247; strigosa, 41

Pratella, 73 Primula angustifolia, 260

Prunulus, 20, 177; galericulatus, 179; viscidipes, 177

Prunus americana, 263; Armeniaca Ansu, 90; donarium, 90; Mume, 90; Persica, 88; salicina, 88, 90

Psalliota, 73

Psathura, 18

Psathyra, 18; albipes, 33; bulbillosa, 22; byssina, 33; cinchonensis, 33; commiscibilis, 19; cubensis, 33; cubispora, 30; diminutiva, 33; epibates, 20; fuliginosa, 33; lateritia, 33; mammillata, 33; mexicana, 33; pallidispora, 33; plana, 33; plumigera, 23; pseudotenera, 25; tigrina,

Psathyrella, 15, 18, 25, 28; arata, 28; cubensis, 25, 27; disseminata, 26; Earlei, 25, 27; grisea, 25, 26; hi-28; mexicana, 25, 26; ascens. minutula, 25, 26; modesta, 29; prona, 26; Stevensonii, 25, 28 Pseudocymopterus montanus.

multifidus, 39

Pseudopeziza Medicaginis, 253

Pseudotsuga, 194, 195, 196, 198; macrocarpa, 197, 198; mucronata, 196; taxifolia, 5, 194, 195, 196, 197

Psilocybe, 15, 17, 18, 29, 33; antillarum, 32; antillarum praelonga, 33; dichroma, 29, 30; fortunata, 33; palmigena, 29; plutonia, 29, 30; orizabensis, 29; subviridis, 30,

Psoralea tenuiflora, 248

Pteris aquilina, 94 Puccinia, Absinthii, 37; aemulans, 37; Andropogonis, 36, 37. 38: Artemisiae, 37; Asteris, 37; Cirsii, 37; Clematidis. 37; Clintonii, 38; conferta, 38; Ellisiana, 37, 38; epi-phylla, 38; Gentianae, 38; Grindeliae, 41; Grossulariae, 38; Heli-anthi, 41; hemispherica, 41; Hieracii, 38; Kuhniae, 38; Menthae, 38; Millefolii, 39; monoica, 198; Muhlenbergiae, 39; poculiformis, 39; Pseudocymopteri, 39; Oxalidis, 41; Saxifragae, 39; Sherardiana, 39; subdecora, 39; substerilis, 39; Taraxaci, 41; tardissima, 39; tuberculans, 39; universalis, 40; vertisepta, 35; violae, 40

Pucciniastrum Agrimoniae, 41; pustulatum, 41

Pycnoporus cinnabarinus, 107; sanguineus, 107

Pyrenophora, 249; chrysospora polaris, 248; comata, 249; Leucelenes,

Pyronema, 277

Pyropolyporus conchatus, 290

Pyroxylophilous fungus, Daldinia vernicosa-a, 277

Pyrus Malus, 90; sinensis, 90

Quamoclidion multiflorum, 246

Quercus, 167; alba, 279, 284; Fendleri, 241, 250, 252

Range of forest fungi, Notes on the altitudinal, 4 Ratibida columnifera, 244

Rhabdospora, 241, 244; dumetorum, 245; gauracea, 262; translucens, 165

Rhizopogon, 222

Rhoads, A. S., Daldinia vernicosa-a pyroxylophilous fungus, 277

Rhododendron albiflorum, 4

Rhus, 250

Rhysotheca Acalyphae, 169; australis, 169; illinoisensis, 169

Rhytisma salicinum, 253

Ribes aureum, 35, 36, 240, 250; Grossularia, 90; inebrians, 35, 36, 251, 252; longiflorum, 36; longifloium, 36; mescalerium, 36; Wolfii, 35

Robillarda Cavarae, 287; Celtidis, 287; mori, 287

Rosa, 242, 243, 244, 250; Fendleri, 37; Maximiliani, 37

Rosellinia, 45, 98; parasitica, 241; pulveracea, 241, 257; Rosarum, 242

Royal palm, A Phyllachora of the, 43 Roystonea regia, 43, 44

Rubus arizonicus, 41

Rudbeckia laciniata, 41, 255

Russula, 93, 95, 267; alutacea, 94; cinerascens, 267; Davisii, 93; disparalis, 94; elatior, 95; fallax, 93; fragiliformis, 93, 96; fragilis, 93; glauca, 93; heterophylla, 93; insignis, 93; integra, 93; Linnaei, 96; magna, 267; paludosa, 95; pectinatoides, 93; perplexa, 96; pulchra, 95, 96; pungens, 267; purpurina, 96; rubrotineta, 95; subvelutina, 96; uncialis, 96; vet-

ernosa, 93 Russula from Massachusetts, New species of, 93

Rusts and smuts collected in New Mexico in 1916, 34

Salix, 36, 90; ballotaeflora, 35; Bebbiana, 253; cordata Watsoni, 36, 252, 263; glandulosa Warburgii, 88; Scouleriana, 36; subcaerulea, 36; Wrightii, 36

Salsola Pestifer, 257 Schizophyllum commune, 7, 100

Schizoxylon insigne, 253 Schmaltzia Bakeri, 250, 252

Sclerotinia parasitica, 86 Scolecosporae, 250, 261

Seaver, Fred J., Photographs and

descriptions of cup-fungi-VII, The genus Underwoodia, 1

Sebacina spongiosa, 98

Senecio amplectens, 248; scopulina, 244; spartioides, 254 acaciae,

Septobasidium, 89; pedicellatum, 90

Septoria gaurina, 261, 262; Helianthi, 261; Oenotherae, 262; smilacina, 262

Sidalcea neomexicana, 255 Sideranthus spinulosus, 39 Sisyrinchium demissum, 259

Sitanion longifolium, 42, 262

Smuts collected in New Mexico in 1916, Rusts and, 34 Solidago Pitcheri, 255

Sparassis radicata, 100

Species of the Collemaceae, A new genus and, 235

Species of fungi from various localities-II, Notes on new, 164 Species, Studies in North American

Peronosporales-VII, New noteworthy, 168 Spermoedia clavus, 251

Sphaeria vernicosa, 283, 284 Sphaeriaceae, 240

Sphaeriales, 239

Sphaeronema, 155, 161, 162; fimbriatum, 155, 156, 160, 162

Sphaeronema fimbriatum, formation in, 155 Sphaeropsidales, 253

Sphaeropsis Diervillae, 164; wistariana, 164

Sphaerotheca Humuli, 240 Sporobolus auriculatus, 248, 259 Stagonospora Chenopodii, 259;

graminella, 259; Humuli-americani, 259; mori, 286

Standley, Paul C., Rusts and smuts collected in New Mexico in 1916,

Stanleyella Wrightii, 259, 263

Stictidaceae, 45 Stilbaceae, 263

Stilbum, 264

Stipa Scribneri, 39; Vaseyi, 39, 42 Strickeria Cercocarpi, 250; rhoina, 250

Stropharia, 16, 68, 70, 71; aeruginosa. 72; ambigua, 101; bermudiensis, 70, 72; caespitosa, 70, 71; cubensis, 70, 72; floccosa, 70, 71; melasperma, 73; semiglobata, 70, 72, 73; stercoraria, 73; troyana, 70

Studies in North American Peronosporales-VII, New and noteworthy species, 168 Studies in the genus Gymnosporangium-III, The origin of the teleutospore, 182

Stypinella Tanakae, 89 Symphoricarpos oreophilus, 242, 252, 253

Tanaka, Tyôzaburô, New Japanese fungi, Notes and translations—IV, 86: V. 285

Taraxacum taraxacum, 41

Tecoma radicans, 165

Teichospora Cercocarpi, 250; obducens, 250; pygmaea, 250; rhoina, 250; rhypodes, 250; stenocarpa, 250 Teleutospore, Studies in the genus Gymnosporangium—III, The origin of the, 182

Thalictrum dasycarpum, 255; Fendleri, 255

Thea sinensis, 88, 90

Thermopsis Pinetorum, 254, 257 Thielavia, 155, 156, 160, 161, 162; basicola, 155, 156, 160

Thyridaria tarda, 45 Thyridium, 240; cingulatum, 240

Trametes carnea, 100; cinnabarina, 107; hispida, 100; serialis, 100; variiformis, 100

ranslations, New Japanese fungi, Notes and, IV, 86; V, 285 Translations-Trifolium Fendleri, 251, 263; nanum, 247; stenolobum, 247

Trimmatostroma Salicis, 263 Triticum aestivum, 39

Tropical North America-Agaricaceae of, VII, 15; VIII, 62 Tsuga heterophylla, 9, 100, 197; mertensiana, 4; taxifolia, 100

Tubercularia vulgaris, 263 Tuberculariaceae, 263 Typha latifolia, 245, 248 Typhula, 57 Tyromyces amorphus, 109

Uncinula flexuosa, 239 Underwoodia, 1; columnaris, 2 Uredinales, 35

Uredinales of Costa Rica based on collections by E. W. D. Holway, III

Uromyces Astragali, 41; Eriogoni, 40: Fabae. 40: Gentianae. 40: 40; Fabae, 40; Gentianae, 40; graminicola, 40; Hedysari-obscuri, 40; intricatus, 40; proeminens, 41; punctatus, 41; Rudbeckiae, 41

Uropyxis sanguinea, 41

Ustilaginales, 41 Ustilago bromivora, 41; Crus-galli, 41; Hordei, 42; hypodytes, 42; levis, 42

Ustulina microspora, 92; mori, 91; zonata, 45, 46

Vaccinium microphyllum, 4 Vagnera stellata, 262 Valeriana, 35 Valsa Paulowniae, 92 Velutina, 233

Venenarius, 101; pantherinoides, 289; pregammatus, 101; umbrinidiscus, TOT

Verbascum Blattaria, 164 Verbena Macdougalii, 250 Veronica, 168; arvensis, 168; peregrina, 168 Vicia americana, 40, 258

Viola pedatifida, 38, 40 Viorna Scottii, 255 Vitis, 90

Weir, James R., Notes on the altitudinal range of forest fungi, 4; Hubert, Ernest E., and, Cultures with Melampsorae on Populus, 194 Wilson, Guy West, Studies in North American Peronosporales - VII, New and noteworthy species, 168

Xanthoxylum, 90 Xylaria Hypoxylon, 45

Yucca baccata, 261; glauca, 257, 259

Zea Mays, 166



